User Guide

Quick-Neuron™ GABAergic - mRNA Kit (Large)

Catalog Number: GA-mRNA-L

Introduction

The Quick-Neuron™ GABAergic - mRNA Kit (Large) facilitates rapid and efficient differentiation of human iPS or ES cells into a population of GABAergic neurons in just 10 days. Our proprietary transcription factor-based stem cell differentiation method uses synthetic mRNAs to produce highly pure populations of neurons without a genetic footprint. Quick-Neuron™ GABAergic differentiated cell cultures display typical neurite outgrowth and express a variety of neuronal markers, such as the pan-neuronal marker tubulin beta 3 class III (TUBB3) and the GABAergic markers parvalbumin (PVALB) and glutamic acid decarboxylase 1 (GAD1). When handled and maintained according to the instructions in this user guide, GABAergic neurons are viable long-term and are suitable for a variety of characterization and neurotoxicity assays.

Scale: The Quick-Neuron™ GABAergic - mRNA Kit (Large) contains a set of reagents for use with a total of 6 wells of a 6-well plate.

Related Products: Quick-Neuron™ GABAergic- mRNA Kit (Small), Catalog Number: GA-mRNA-S
Quick-Neuron™ GABAergic- Human iPSC-derived Neurons, Catalog Number: GA-mRNA-CW
Quick-Neuron™ GABAergic- Maintenance Medium, Catalog Number: GA-MM

Contents

Upon receipt, store the reagents at the temperatures indicated in the table below. All reagents are shipped on dry ice.

Contents	Volume	Storage	Thaw
QNG-mRNA-P	4 x 33 µl	-80°C	On ice
Component N	4 x 840 µl	-20°C or -80°C	On ice or 4°C
Component P	2 x 50 µl	-20°C or -80°C	Room temperature
Component G1	3 x 20 µl	-20°C or -80°C	On ice or 4°C
Component G2	60 µl	-20°C or -80°C	On ice or 4°C

Condition of Use

This product is for research use only. It is not approved for use in humans or for therapeutic or diagnostic use.

Technical Support

For technical support please refer to the FAQ on our website.
You may also contact us at cs@elixirgensci.com or call +1 (443) 869-5420 (M-F 9am-5pm EST).

Required Consumables

Item	Vendor	Catalog Number
6-well tissue-culture-treated polystyrene plate (e.g., Corning Costar Flat Bottom Cell Culture Plates)	Fisher Scientific	07-200-80
(Optional) 24-well tissue-culture-treated polystyrene plate (e.g., Corning Costar Flat Bottom Cell Culture Plates)	Fisher Scientific	07-200-740
(Optional) 96-well tissue-culture-treated polystyrene plate (e.g., Thermo Scientific™ 96 Well Black/Clear Bottom Plate)	Fisher Scientific	12-566-70
Lipofectamine MessengerMAX	ThermoFisher	LMRNA001
Opti-MEM I Reduced Serum Medium	ThermoFisher	31985062
DMEM/F12	ThermoFisher	21331020
Neurobasal Medium	ThermoFisher	21103049
GlutaMAX	ThermoFisher	35050061
Penicillin-Streptomycin	ThermoFisher	15140122
StemFit Basic04 Complete Type, or StemFit AK02N, or StemFlex Medium	Elixirgen Scientific TaKaRa ThermoFisher	ASB04-C, or AK02N, or A3349401
iMatrix-511 silk	Elixirgen Scientific	NI511S
TrypLE Select Enzyme (1X)	ThermoFisher	12563011
0.02% EDTA in DPBS	Sigma-Aldrich	P4957-50ML
0.01% Poly-L-Ornithine	Sigma-Aldrich	P4957-50ML
Extracellular Matrix such as** - Laminin Mouse Protein, Natural, or - Geltrex Basement Membrane Matrix	ThermoFisher	23017015 or A15696-01
Phosphate-buffered saline (without Ca^{++} Mg^{++})*	ThermoFisher	20012050
ROCK inhibitor Y27632	Selleckchem	S1049
Dimethyl sulfoxide (DMSO)	Sigma-Aldrich	D2650
Puromycin (10 mg/ml)	InvivoGen	ant-pr-1

* PBS should be used at room temperature unless otherwise specified.

Source hPSC Culture Conditions

The Quick-Neuron™ GABAergic - mRNA Kit (Large) gives the best differentiation results when source human pluripotent stem cells (hPSCs) have been maintained in StemFit® Basic04, StemFit® AK02N, StemFlex™ Medium, or other similar culture media which enable the maintenance of cultures by single-cell passaging. This protocol also assumes that the source hPSCs are cultured in a 35-mm culture dish or wells of a 6-well plate. If iMatrix-511 silk is routinely used as a coating substrate, prepare culture dishes or wells precoated with 0.25 µg/cm^2 iMatrix-511 silk diluted in 2 ml chilled PBS per well or dish for this kit.

- The protocols and reagents for StemFit® Basic04, and iMatrix-511 silk culture conditions are available at Elixirgen Scientific (Catalog Numbers: ASB04-C, NI511S).
- Differentiation should not be performed until the cells are at least 14 days post-thaw.

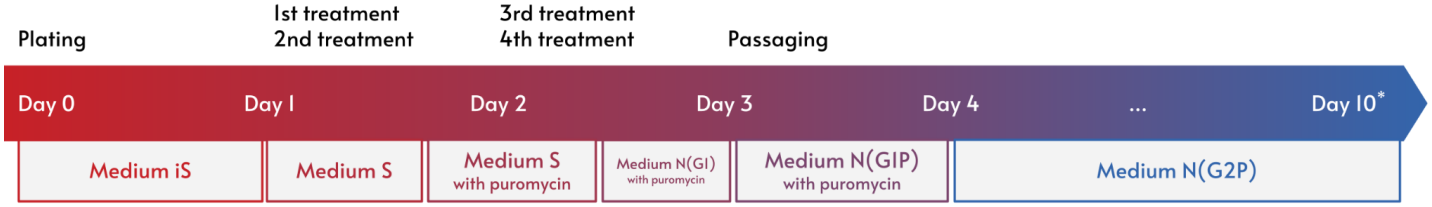
- We recommend preparing a minimum of 3.6 x 10^6 viable hPSC for use with this kit. This is usually obtained by using 2-3 wells of a 6-well plate at 50-70% confluency.
- For optimal differentiation, hPSC confluency should be around 50% to 70%. Do not use wells more than 90% confluent.

Drug Selection

Users should perform a puromycin kill curve for their cells to determine the minimum concentration required to kill all non-treated cells within ~60 hours. Based on Elixirgen Scientific's internal tests, the appropriate concentration ranges between 0.5 and 2 µg/ml. We recommend maintaining 2 wells of untransfected iPSC (with standard StemFit conditions), alongside the transfected wells, until after puromycin selection is performed. Treat 1 of those wells with puromycin at your selected concentration so as to confirm that the puromycin is effective at killing the untransfected cells in your experiment.

Workflow

Note: This protocol assumes that Day 0 is a Monday.



* From Day 10, users may maintain differentiated neurons in the medium best suited for their needs, though we recommend Quick-Neuron™ GABAergic - Maintenance Medium, Catalog Number: GA-MM.

Preparation

Important Note! For the best possible delivery of QNG-mRNA-P into cells, we recommend Lipofectamine MessengerMax. If users prefer another transfection reagent, please make sure that the reagent provides a transfection efficiency of ≥80% prior to using this kit. QNG-mRNA-P mixed with Lipofectamine MessengerMax must be immediately applied to cultures and cannot be stored.

10 mM ROCK inhibitor Y27632 (iROCK)

1. Dissolve 10 mg ROCK inhibitor Y27632 in 3.12 ml DMSO.
2. Make aliquots of a convenient volume (e.g., 100 µl).
3. This solution, hereafter referred to as iROCK, can be stored at -20°C.

StemFit Basic04 Complete Type (Medium S)*

1. Thaw StemFit Basic04 Complete Type bottle overnight or multiple nights at 4°C.
2. Make aliquots of a convenient volume (e.g., 40 ml).
3. This solution, hereafter referred to as Medium S, can be stored at -80°C. Once thawed, Medium S should be stored at 4°C for up to 2 weeks.
    - After thawing users may choose to add Penicillin-Streptomycin at a 1:200 dilution (e.g., 200 µl in 40 ml of Medium S) before using Medium S.

*Medium S can be substituted with StemFit AK02N or StemFlex.

0.5X TrypLE Select with EDTA (Solution D1)

1. Mix 1 ml TrypLE Select Enzyme (1X) with 1 ml 0.02% EDTA in DPBS.
2. This mixture, hereafter referred to as Solution D1, can be stored at 4°C for 2 weeks.

0.002% Poly-L-Ornithine solution (ornithine)

1. Take 2 ml 0.01% Poly-L-Ornithine solution and mix it with 8 ml PBS.
2. The 0.002% Poly-L-Ornithine solution, hereafter referred to as ornithine, can be stored at 4°C for up to 2 weeks.

1 mg/ml laminin stock solution (laminin)

1. Thaw Laminin Mouse Protein, Natural and chill PBS at 4°C or on ice.
2. Mix the Laminin Mouse Protein, Natural and PBS to make the 1 mg/ml stock solution, hereafter referred to as laminin.
    - Laminin concentration varies by lot, so use the number specified on the vial or CoA when making your calculations.
3. Make aliquots of a convenient volume (e.g., 90 µl) and store at -20°C.

Medium N

1. Prepare Medium N using the reagents listed in the table below.
    - Thaw Component N for 20-30 minutes at the temperature indicated in the “Contents” table on page 1.
    - Warm all other reagents at room temperature for 20-30 minutes.
    - Tap Component N tubes 3 times and then briefly spin all tubes down before use.
    - Keep Medium N, and any subsequent media made with it, protected from light.
    - Store Medium N for up to 2 weeks at 4°C.
    - Leftover Components N can be discarded or saved at 4°C for up to two weeks.

Reagents	Volume
DMEM/F12	41.5 ml
Neurobasal	41.5 ml
GlutaMAX	435 µl
Penicillin-Streptomycin (10000 units/ml; 100x)	870 µl
Component N	2.7 ml

Day -3

~1 hour

Note: This protocol assumes that Day 0 is a Monday so Day -3 is Friday.

Plate Preparation

1. Prepare diluted Coating Material A by mixing together the following components in a 15 ml conical tube.
    - Thaw Coating Material A for 20-30 minutes at the temperature indicated in the “Contents” table on page 1.
    - Keep Coating Material A on ice.
    - Make sure chilled PBS is used for this mixture.

Reagents	Volume
Coating Material A	11.6 µl
Chilled PBS	3.5 ml

2. Add 400 µl diluted Coating Material A to each new well of 8 wells.
3. Incubate the plate at 4°C.

Note: For best results we recommend precoating the plate 1 day or up to a week before use and keeping at 4°C. Alternatively plates can be precoated on Day 0 and placed at 37°C for at least 2 hours before use.

Day 0

~4 hours

Note: This protocol assumes that Day 0 is a Monday and that user's hPSCs were already used with a small size kit (Catalog number: GA-mRNA-S) so that users are familiar with the experimental process and have optimized conditions for their particular cells.

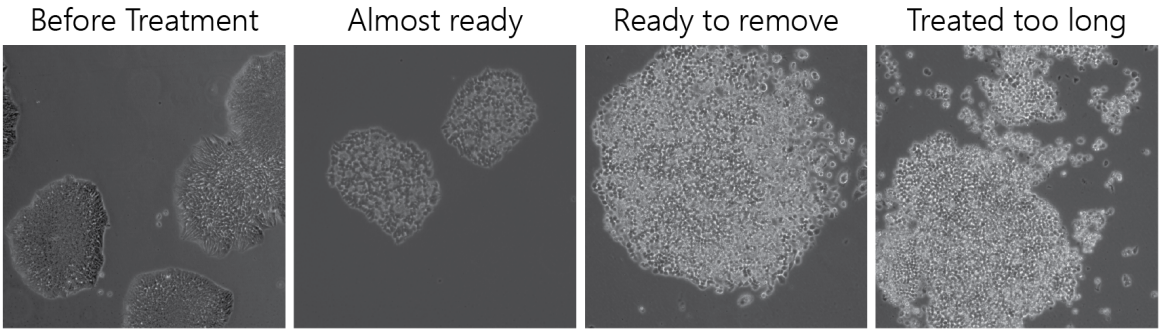
Plating

IMPORTANT! Source hPSC wells should be no more than 50-70% confluent thus requiring a minimum of 2 wells to begin differentiation.

1. Determine the number of wells required to get 3.6 x 10^6 cells from the source hPSC 6-well plate.
   NOTE: Cells will be plated in a new 6-well plate at 3 densities (0.5 x 10^6 cells, 0.55 x 10^6 cells, and 0.6 x 10^6 cells), with 2 wells per density.
2. Prepare Medium iS by mixing together the following components in a 15 ml conical tube.
   - Warm Medium S, iROCK, and Solution D1 at room temperature for at least 1 hour protected from light.
   - The rest of Medium S should be stored at 4°C for later use.

	Required medium volume based on # of wells of a 6-well plate	
Reagents	**2 wells**	**3 wells**
Medium S	12.7 ml	15.5 ml
iROCK	12.7 µl	15.5 µl

3. Aspirate old medium from hPSC culture and add 1.5 ml of Medium iS to each well.
4. Incubate the culture at 37°C, 5% CO_2 for 1 hour before harvesting cells.
   - This is to decrease cell death on Day 1 and minimize the loss of cells.
5. Aspirate old medium from hPSC culture and add 2 ml PBS to each well being harvested.
6. Rock the plate 3 times, aspirate PBS from the culture, and add 300 µl of the cell dissociation reagent Solution D1.
   - Keep the rest of Solution D1 at 4°C for use on Day 3.
7. Incubate the culture plate at 37°C, 5% CO_2 for 5 minutes. If all the cells are not rounded under a microscope, continue to incubate at 37°C, 5% CO_2 in 1-2 minute increments (see images below).



8. Carefully pipet out Solution D1 from the culture and add 1 ml Medium iS to the well.
   - Follow Steps 8-10 one well at a time if multiple wells are used.
9. Disperse the medium over the bottom surface of the well by pipetting 8-15 times to detach cells.
10. Using the same pipet tip, collect the cell suspension in a 15 ml tube.
11. Count cells and determine viability.
12. Take out the volume of the cell suspension needed for 2 wells of each cell density, according to the note in step 1, and include an extra 10% as a buffer. Place each in a new tube labeled with the corresponding density.

13. Bring the volume of the cell suspension in each tube up to 2.2 ml with Medium iS.
    - If the volume in the tube exceeds 2.2 ml, centrifuge the required volume of cell suspension at 200 x g for 4 minutes, remove the supernatant, and resuspend the pellet in 2.2 ml Medium iS.
14. Aspirate diluted iMatrix-511 silk from each newly coated well and add 1 ml cell suspension to each well.
    - Most of the diluted iMatrix-511 silk should be aspirated but not completely to prevent the coated well from drying before adding the cell suspension. The cell suspension should be added to the well immediately after the diluted iMatrix-511 silk is removed. Handle one well after another.
15. Leave the plate flat at room temperature for 10 minutes.
16. Incubate the culture plate at 37°C, 5% CO_2 overnight.

Day 1

~8 hours

IMPORTANT! Observe all wells under a microscope and confirm that all 6 wells show 50-70% confluency for transfections with QNG-mRNA-P. If there are any wells that do not fall within the range of confluence, do not use them.

First Treatment

1. Thaw 1 vial of QNG-mRNA-P on ice for 30 minutes and warm Opti-MEM and Medium S at room temperature for 20-30 minutes.
2. Prepare QNG by the following steps:
    - Tap the QNG-mRNA-P tube 3 times and then briefly spin it down before use
    - Prepare a 15 ml tube and a 1.5 ml tube with 825 µl Opti-MEM each. Label the 15 ml tube “Mix 1” and the 1.5 ml tube “Mix 2”.
    - Add 16.5 µl Lipofectamine MessengerMax (LMM) to the Mix 1 tube and mix by brief vortexing. Leave it at room temperature for 10 minutes (Mix 1). Keep the rest of LMM at 4°C for later treatments.
    - **IMPORTANT!** Immediately before 10 minutes pass (i.e., around 8 minutes), add the entire contents of the QNG-mRNA-P vial to the other 1.5 ml tube with Opti-MEM (Mix 2). Mix by tapping 5 times. Do not vortex.
    - 10 minutes after mixing LMM with Opti-MEM, add Mix 2 into Mix 1, and pipet up and down 8-10 times. This mixture is called QNG. Leave QNG at room temperature for 5 minutes and no longer.

Mix 1 Reagents	Volume
Opti-MEM	825 µl
LMM	16.5 µl

Mix 2 Reagents	Volume
Opti-MEM	825 µl
QNG-mRNA-P	~33 µl

3. Add 6.6 ml Medium S to QNG and pipet up and down 2-3 times to mix.
4. Working with up to 2 wells at a time, aspirate the old medium out and add 1.25 ml of QNG mixture to each well. Repeat until QNG mixture has been added to all wells.
5. Incubate the culture plate at 37°C, 5% CO_2 for 2.5 hours.

Second Treatment

1. Pipet out the medium from each well and add 1 ml Medium S.
2. Incubate the culture plate at 37°C, 5% CO_2 for 2 hours.
    - Put 1 vial of QNG-mRNA-P on ice when the incubation above reaches 1.5 hours and leave it for 30 minutes.
3. Repeat Steps 2-5 of the previous “First Treatment” section.

Medium Change and Drug Selection

1. Transfer 10 ml Medium S into a tube and add puromycin to it at the predetermined optimal concentration (see earlier section on “Drug Selection”).
2. Pipet out the medium from each well and add 1.5 ml Medium S with puromycin.
3. Incubate the culture plate at 37°C, 5% CO_2 overnight.

Day 2

~8 hours

IMPORTANT! Observe the QNG-treated cultures to make sure that they are reaching confluency (≥90%). If the cultures are <50% confluent and show signs of cell death (e.g., many floating cells), users should skip the third and fourth treatments on Day 2 and proceed directly to the steps described in Day 3. The protocol will then be accelerated by one day.

Third Treatment

1. Thaw 1 vial of QNG-mRNA-P on ice for 30 minutes and warm Opti-MEM and Medium S at room temperature for 20-30 minutes.
2. Repeat Steps 2-5 of the previous "First Treatment" section.

Fourth Treatment

1. Pipet out the medium from each well and add 1 ml Medium S.
2. Incubate the culture plate at 37°C, 5% CO_2 for 2 hours.
    - Put 1 vial of QNG-mRNA-P on ice when the incubation above reaches 1.5 hours and leave it for 30 minutes.
3. Repeat Steps 2-5 of the previous "First Treatment" section.

Medium Change and Drug Selection

1. Prepare Medium N(G1) using the volumes indicated in the table below.
    - Thaw Component G1 for 20-30 minutes at the temperature indicated in the "Contents" table on page 1.
    - Warm Medium N at room temperature for 20-30 minutes.
    - Tap the Component G1 tube 3 times and then briefly spin it down before use
    - Store Medium N(G1) for up to 2 weeks at 4°C.
    - Store the remaining Medium N and thawed Component G1 at 4°C.

Reagents	Volume
Medium N	10 ml
Component G1	20 µl

2. Add puromycin to Medium N(G1) at the predetermined optimal concentration (see earlier section on "Drug Selection").
    - If more than 90% of the cells show resistance to puromycin at the concentration used on Day 1, consider increasing its concentration.
3. Pipet out the medium from each well and add 1.5 ml Medium N(G1) with puromycin.
4. Incubate the culture plate at 37°C, 5% CO_2 overnight.

Day 3

4-6 hours

New Plate Preparation

IMPORTANT! This kit can accommodate replating to all wells of either a 6-well, a 24-well, or a 96-well plate. Refer to the tables for the recommended volumes. Please note that the volumes are per well in Table A and per plate in Table B.

1. Vortex ornithine briefly and centrifuge it at a maximum speed for 1 minute.
2. Add ornithine to each well of a new plate in the volume specified in Table A.
3. Incubate the plate at 37°C, 5% CO_2 for at least 2 hours (or at 4°C overnight one day before plating).
4. Thaw laminin and chill specified amounts of PBS on ice for 20-30 minutes.
5. Add laminin to chilled PBS in the volume specified in Table B. Mix well.
    - All PBS washes should be done dropwise and with room temperature PBS. Chilled PBS is only for the coating step.
6. Aspirate the supernatant from each well and add PBS in the volume specified in Table A.

7. Repeat Step 6.
8. Aspirate PBS from each well and add diluted laminin according to Table A.
9. Incubate the plate at 37°C, 5% CO_2 for at least 2 hours.

Table A. Recommended volumes per well for different plate formats.

Reagents	Recommended volume per well		
	6-well plate	24-well plate	96-well plate
Ornithine	1.5 ml	300 µl	50 µl
PBS	2 ml	500 µl	100 µl
Diluted laminin	1.5 ml	300 µl	50 µl
Medium N(G1P) with puromycin	500 µl	200 µl	35 µl

Table B. Recommended volumes per plate for different plate formats.

Reagents		Recommended volume per plate		
		6-well plate	24-well plate	96-well plate
Diluted laminin	Laminin	100 µl	80 µl	53 µl
	Chilled PBS	10 ml	8 ml	5.3 ml
Medium N(P)	Medium N	68 ml	75 ml	68 ml
	Component P	68 µl	75 µl	68 µl

Media Preparation

1. While the plate is incubating, prepare Medium N(P) using the volumes indicated in Table B above.
    - Thaw Component P for 20-30 minutes at the temperature indicated in the "Contents" table on page 1.
    - Warm Medium N at room temperature for 20-30 minutes.
    - Tap each Component tube 3 times and then briefly spin all tubes down before use.
    - Keep the rest of Medium N at 4°C for later use.
2. Prepare Medium N(G1P) using the volumes indicated in the table below.
    - Thaw Component G1 for 20-30 minutes at the temperatures indicated in the "Contents" table on page 1.
    - Tap each Component tube 3 times and then briefly spin all tubes down before use.
    - Keep the rest of Medium N(P) at 4°C for later use.

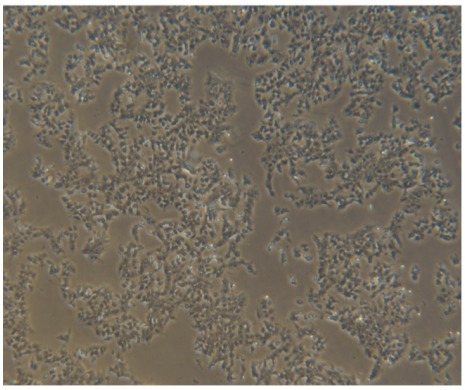
Reagents	Volume
Medium N(P)	12.5 ml
Component G1	24 µl

3. Add puromycin to the Medium N(G1P) at the predetermined optimal concentration (see earlier section on "Drug Selection").
4. After the laminin incubation, aspirate most, but not all, of the supernatant from each well of the new plate and add PBS in the amount specified in Table A above. Add the PBS dropwise to each well.
5. Aspirate most, but not all of the PBS and add Medium N(G1P) with puromycin in the volume specified in Table A above.
6. Incubate the plate at 37°C, 5% CO_2 until cells are ready for plating.

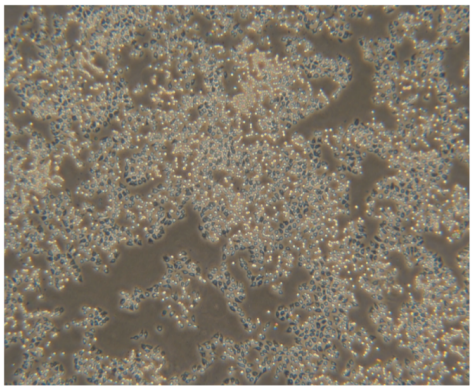
Passaging Cells

IMPORTANT! For the following steps, gently pipet and add solutions. Differentiating cells are delicate and should be handled with great care. **Perform these steps one well at a time.** Refer to the images below to successfully manage cell treatment and dissociation.

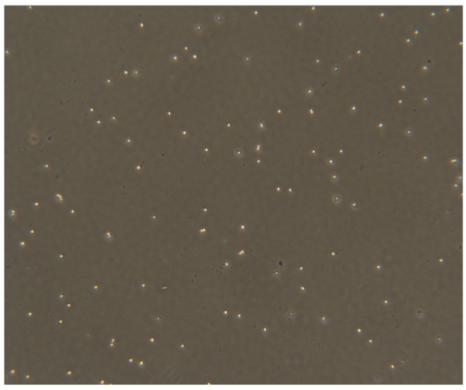
Before Solution D1 treatment



During Solution D1 treatment



After Dissociation



1. Make sure that Solution D1 is at room temperature for at least 1 hour before use.
2. Pipet out the old medium from one well and add 1 ml PBS to the well.
3. Pipet out the PBS from the well and add 300 µl Solution D1.
4. Rock the plate 3 times to spread the Solution D1 evenly.
5. Incubate the cultures at 37°C, 5% CO_2 for 3 minutes.
6. Carefully pipet out Solution D1 from the well and add 750 µl Medium N(G1P) with puromycin along the wall of the well.
7. Disperse the medium quickly over the bottom surface of the well by pipetting 6-8 times to detach cells.
8. Observe cells and/or cell aggregates floating in the well under a microscope. It is normal that 10-20% of cells remain attached to the well bottom after pipetting. These clusters of cells are not supposed to be lifted. Do not attempt to detach all of the cells remaining on the well bottom.
9. Collect 750 µl cell suspension from the well and transfer to a tube.
10. Repeat steps 2-9 for the rest of the wells.
11. Gently pipet the cell suspension up and down up to 5 times to break the cell aggregates. Excessive pipetting can damage the already-suspended neuronal cells.
12. Count cells and determine viability.
13. Prepare specified amounts of a 1 x 10^6 live cells/ml cell suspension using Medium N(G1P)with puromycin based on the table below.
14. Add cell suspension to the center of each well. Since each well already has Medium N(G1P) with puromycin, the total volume of the medium in each well is indicated in the table below.

	Recommended Amounts		
	6-well plate	24-well plate	96-well plate
Viable cells/well	5 x 10^5 cells	1 x 10^5 cells	1.5 x 10^4 cells
Req vol of cell suspension (1 x 10^6 viable cells/ml) • (Vol of cell suspension/well x # of wells) + 10% extra	3.3 ml	2.64 ml	1.6 ml
Volume of cell suspension/well	500 µl	100 µl	15 µl
Total volume/well • Medium N(G1P) with puromycin + cell suspension	1 ml	300 µl	50 µl

15. Incubate the culture plate at 37°C, 5% CO_2 overnight.

Day 4

< 2 hours

Medium Change

IMPORTANT! It is optional, but recommended, to include the PBS wash if cell death/floating cells are observed.

1. Prepare Medium N(G2P) using the volumes indicated in the table below.
    - Thaw Component G2 for 20-30 minutes at the temperature indicated in the “Contents” table on page 1.
    - Warm Medium N(P) at room temperature for 20-30 minutes.
    - Tap the Component G2 tube 3 times and then briefly spin it down before use
    - Store Medium N(G2P) for up to 2 weeks at 4°C.

	Required volume per well		
Reagents	**6-well plate**	**24-well plate**	**96-well plate**
Medium N(P)	50 ml	56 ml	50 ml
Component G2	50 µl	56 µl	50 µl

2. Pipet out the old medium from each well and * add Medium N(G2P) according to the table below.
   *(Optional) Slowly add PBS, in the amount specified in the table, alongside the wall of each well to avoid lifting attached cells. Gently pipet out PBS before adding Medium N(G2P).

	Required volume per well		
Reagents	**6-well plate**	**24-well plate**	**96-well plate**
(Optional) PBS	1 ml	500 µl	100 µl
Medium N(G2P)	2 ml	800 µl	150 µl

3. Incubate the culture plate at 37°C, 5% CO_2 for 2 days.

Day 7

< 1 hour

Medium Change

IMPORTANT! It is optional, but recommended, to include the PBS wash if cell death/floating cells are observed.

1. Warm Medium N(G2P) at room temperature for 20-30 minutes.
2. Pipet out the old medium from each well and * add Medium N(G2P) according to the table below.
   *(Optional) Slowly add ml PBS, in the amount specified in the table, alongside the wall of each well to avoid lifting attached cells. Gently pipet out PBS before adding Medium N(G2P).

	Required volume per well		
Reagents	**6-well plate**	**24-well plate**	**96-well plate**
(Optional) PBS	1 ml	500 µl	100 µl
Medium N(G2P)	4 ml	1 ml	250 µl

3. Incubate the culture plate at 37°C, 5% CO_2.

Day 9

Medium Change

1. Warm Medium N(G2P) at room temperature for 20-30 minutes.
2. Pipet out half the volume of old medium from each well (see table on Day 7 for original volume) and replace with an equal volume of fresh Medium N(G2P) according to the table below.

	Required volume per well		
Reagents	**6-well plate**	**24-well plate**	**96-well plate**
Medium N(G2P)	1.5 ml	400 µl	75 µl

3. Incubate the culture plate at 37°C, 5% CO_2.

Day 10

Assay or Continuous Maturation

- Differentiated neurons can be observed on Day 5. For more mature neurons, we recommend culturing cells until Day 10. From Day 10, users may maintain differentiated neurons in the medium best suited for their needs, though we recommend Quick-Neuron™ GABAergic Maintenance Medium, Catalog Number: GA-MM.
- Differentiation into GABAergic neurons after using Quick-Neuron™ GABAergic - mRNA Kit can be confirmed with the markers TUBB3, PVALB, and GAD1.

Appendix A

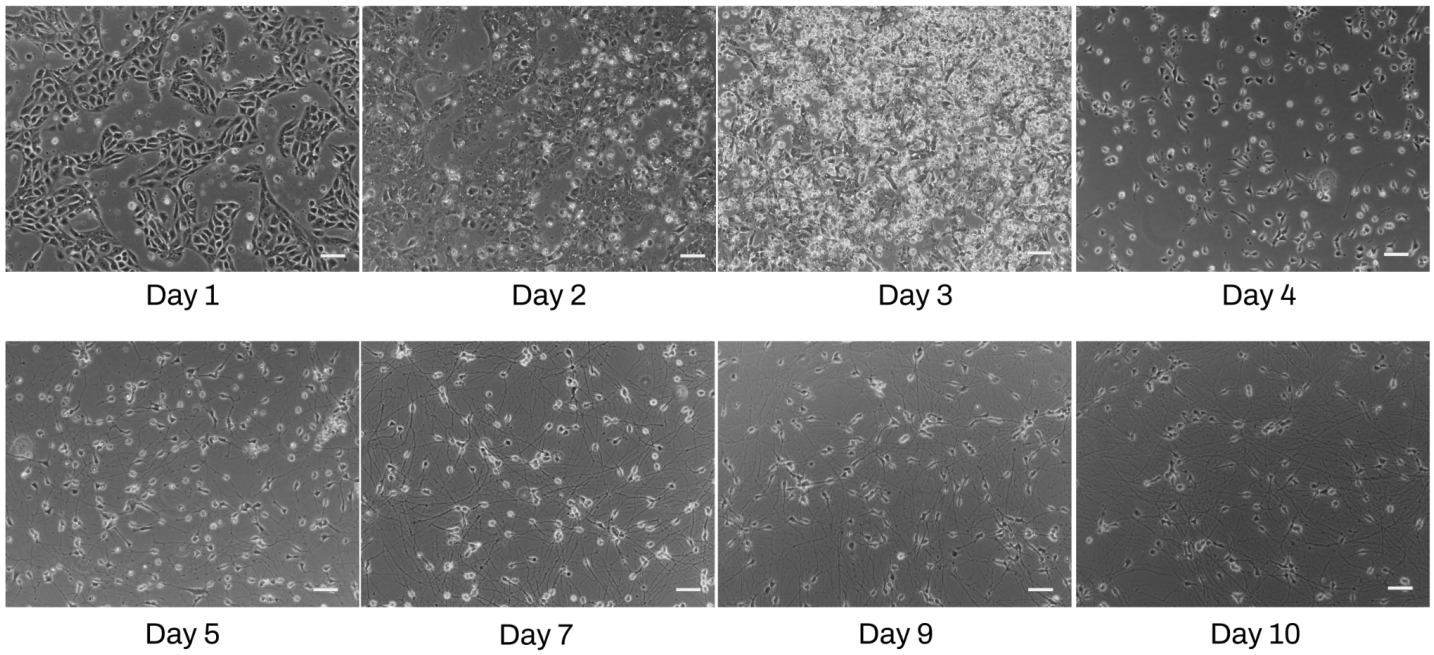


Figure 1. Representative phase contrast images of Quick-Neuron™ GABAergic - mRNA Kit cell cultures on days 1, 2, 3, 4, 5, 7, 9 and 10 post-differentiation (scale bar = 100 µm).

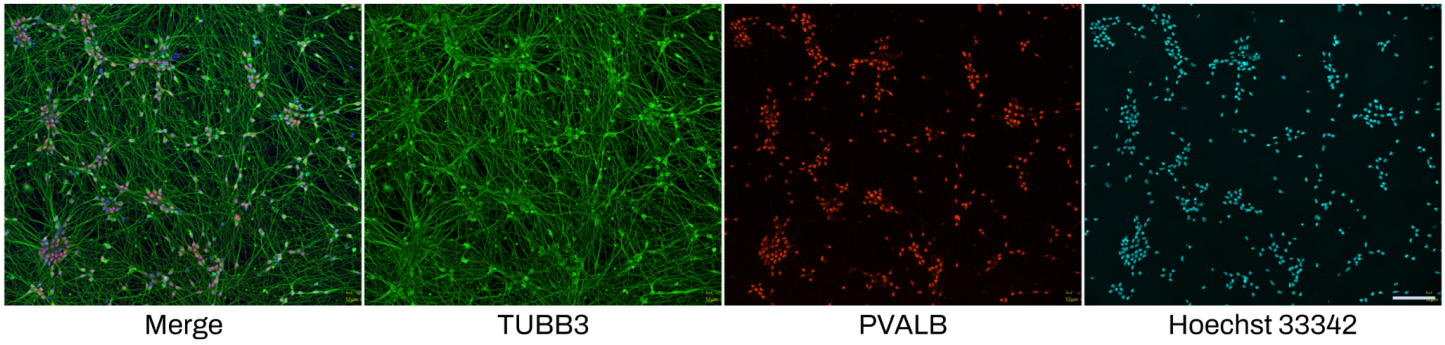


Figure 2. Immunofluorescent staining of Quick-Neuron™ GABAergic - mRNA Kit cell cultures shows typical neurite growth and expression of the pan-neuronal marker TUBB3 and the GABAergic neuron-specific marker PVALB on day 10 post-differentiation (scale bar = 100 µm). Staining conditions: Anti-β-III tubulin monoclonal antibody (R&D Systems, Catalog Number: MAB1195, 1:250 dilution) in combination with a secondary antibody (Invitrogen, Catalog Number: A32723 Goat anti-Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody, AlexaFluor Plus 488, 1:500 dilution). Anti-PVALB primary antibody (Novus Biologicals, Catalog Number: NB120-11427 , 1:1000 dilution) in combination with a secondary antibody (Invitrogen, Catalog Number: A11037 Goat anti-Rabbit IgG (H+L) Highly Cross-Adsorbed Secondary Antibody, Alexa Fluor 594, 1:500 dilution). Nuclei were counterstained with Hoechst 33342.

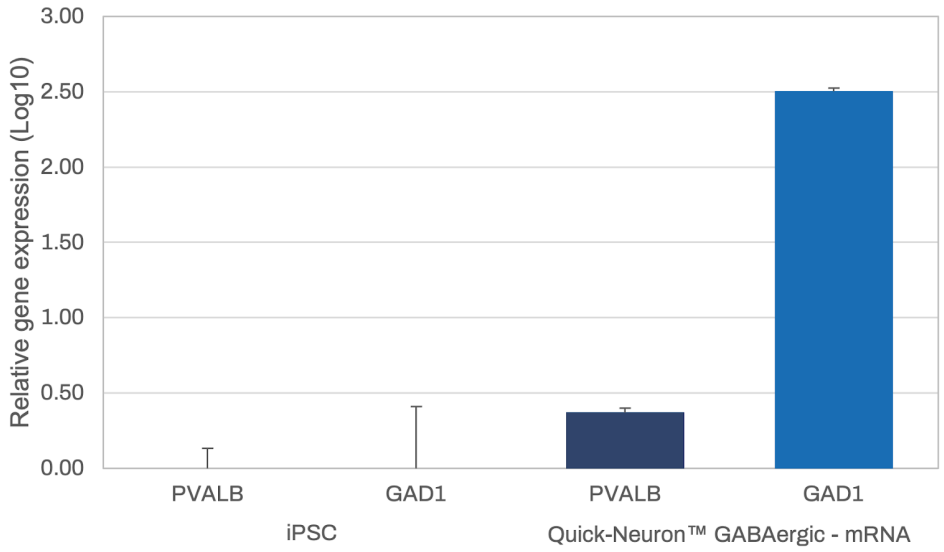


Figure 3. Real-time quantitative PCR analysis of expression levels of GABAergic neuron-associated genes *PVALB* and *GAD1* were examined for Quick-Neuron™ - GABAergic - mRNA Culture on day 10. The relative gene expression is normalized to phosphoglycerate kinase 1 (*PGK1*), and then calculated as a fold induction relative to undifferentiated hPSCs as a control. Error bars show standard deviation. Primers used are listed in Table 1.

Table 1. List of PCR primers used in Figure 3

Gene	Forward Primer	Reverse Primer	Primer Concentration
PVALB	TCGACCACAAAAAGTTCTTC	TTTAGGATGAATCCCAGCTC	250 nM
GAD1	GTCGAGGACTCTGGACAGTA	GGAAGCAGATCTCTAGCAAA	250 nM
PGK1	GTATGCTGAGGCTGTCACTCG	CCTTCCAGGAGCTCCAAACTGG	250 nM